



## Peptidyl $\alpha$ -Keto Thiazole as Potent Thrombin Inhibitors

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**Abstract:** We report the synthesis and evaluation of  $\alpha$ -keto thiazole derivatives such as D-Phe-Pro-Arg-thiazole **9** as a novel type of thrombin inhibitor. Tripeptidyl  $\alpha$ -keto thiazole **9** exhibited the inhibitory activity of thrombin at nanomolar levels and showed a more potent prolongation effect on clotting time than argatroban at a dose of 3 mg/kg intravenously. © 1997 Elsevier Science Ltd. All rights reserved.

### Introduction

Thrombin (EC 3.4.21.5) is a serine protease and the key enzyme in the blood coagulation cascade.<sup>1</sup> Thrombin catalyzes the conversion of fibrinogen to fibrin, which then polymerizes to form a hemostatic plug. Thus, there has been considerable interest in research for thrombin inhibitors as anticoagulant agents.<sup>2</sup>

An approach to the design of the serine protease inhibitor has been the replacement of the scissile amide bond by an electron-withdrawing carbonyl group.<sup>3</sup> Edwards *et al.* originally reported that  $\alpha$ -keto benzoxazole derivative was a mechanism-based elastase inhibitor in which the nitrogen atom of the benzoxazole interacted with the histidine residue of the catalytic triad in serine protease.<sup>4</sup> Recently, we elucidated that  $\alpha$ -keto heterocyclic compounds possessed potent inhibitory activity for prolyl endopeptidase (PEP).<sup>5</sup> In the course of our PEP inhibitors study, we substituted the amino acid residue in the P<sub>1</sub> position of phenylbutanoyl-Pro-A-thiazole **1-5** (A: Pro, Ala, Val, Lys, and Arg) (Table 1). The basic residue of Arg or Lys was indispensable for the inhibitory activity of thrombin, but not PEP.

**Table 1. Structure and inhibitory potencies of  $\alpha$ -keto thiazole derivatives.**

**Ph(CH<sub>2</sub>)<sub>3</sub>CO-Pro-A-thiazole**

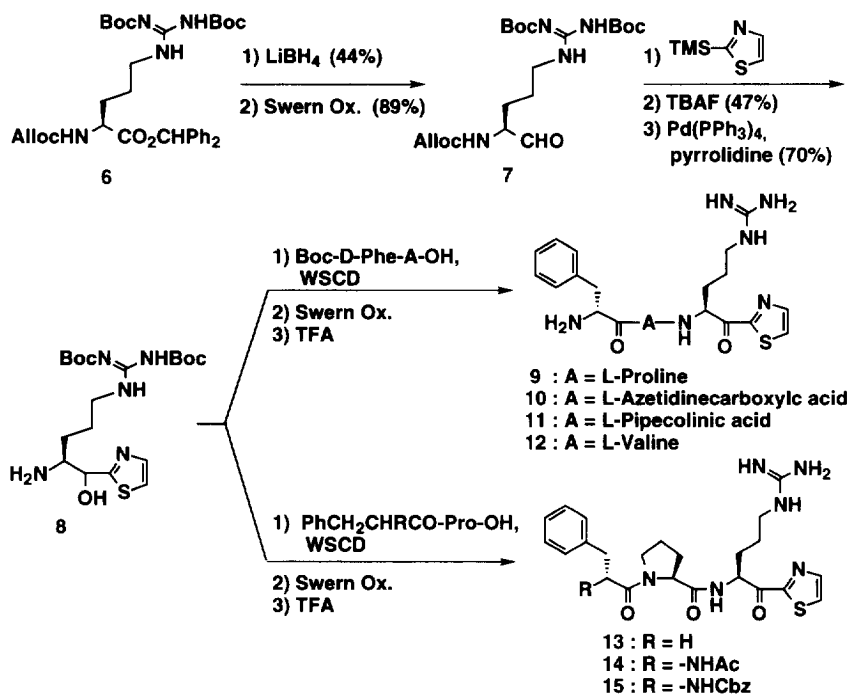
No	A (amino acid)	Inhibitory Activity IC <sub>50</sub> (μM)			
		PEP	Elastase	Thrombin	Trypsin
1	Proline	0.0044	>1000	>1000	>1000
2	Alanine	0.0050	>1000	>1000	>1000
3	Valine	7.7	>1000	>1000	>1000
4	Lysine	7.8	Not determined	46	2.4
5	Arginine	28	>1000	2.0	0.23

These results also indicated that  $\alpha$ -keto heterocyclic compound was useful for research on other serine protease inhibitors. Tripeptide aldehyde and chloromethyl ketone analogs of D-Phe-Pro-Arg sequence are reported to be high effective and reversible inhibitors of thrombin.<sup>2</sup> Considering these factors for inhibitor design, we examined the structure-activity relationship of the tripeptide  $\alpha$ -keto thiazole compounds in the P<sub>1</sub>, P<sub>2</sub> and P<sub>3</sub> positions.

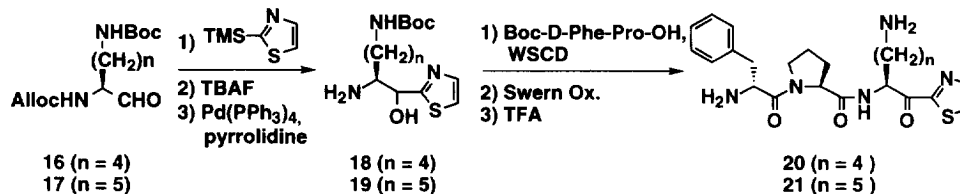
**Chemistry:** We synthesized a series of tripeptide arginine, lysine, and homolysine derivatives with  $\alpha$ -keto thiazole moiety. All compounds were obtained in a convergent strategy as shown in Scheme I. The arginine aldehyde **7** was prepared by the reduction of N-Allyloxycarbonyl(Alloc) arginine ester **6** followed by Swern oxidation.<sup>6</sup> The reaction of aldehyde **7** with 2-(trimethylsilyl)thiazole gave the 2-thiazole derivative, which was converted to amino alcohol **8** by deprotection of the Alloc group. Condensation of the amino alcohol **8** with Boc-D-Phe-Pro-OH followed by Swern oxidation gave the tripeptide. The desired  $\alpha$ -keto thiazole compound **9** was prepared by the TFA deprotection of the Boc protective group. In a similar manner, the corresponding arginine derivatives **10–15**, lysine derivative **20**, and homolysine derivative<sup>7</sup> **21** were synthesized.<sup>6</sup>

### Scheme I

#### (1) Preparation of arginine derivatives



#### (2) Preparation of lysine and homolysine derivatives



**Enzyme assay:** Human thrombin was purchased from Sigma Chemical Company Ltd. Thrombin and trypsin assays were performed as described by Kawabata.<sup>8</sup> PEP and elastase assays were carried out as described by Walter<sup>9</sup> and Bieth<sup>10</sup>, respectively. Urokinase, plasmin, and plasma kallikrein assays were performed by the method used by Morita.<sup>11</sup> Experiments were conducted in 96-well plates, and the rates of hydrolysis were measured fluorometrically with excitation at 380 nm and emission at 440 nm or spectrophotometrically at 405 nm.

**Ex vivo anticoagulant studies:** Five minutes after intravenous administration of test compound **9** and argatroban<sup>12</sup>, the prolonged activated partial thromboplastin time (APTT) and prothrombin time (PT) were determined by the method of Takamiya<sup>13</sup> and Suzuki<sup>14</sup>, respectively.

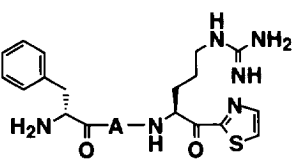
**Results & Discussion:** Replacement of the arginine in **9** with a lysine or homolysine residue at the P<sub>1</sub> position resulted in a marked decrease in the inhibitory potency of thrombin, although the thrombin to trypsin ratios remained unchanged (Table 2). Unexpectedly, the homolysine derivative **21**, which possesses carbon chains of the same length as arginine, was less potent than lysine derivative **20**. These results showed that the arginine in the P<sub>1</sub> position was most suitable for the thrombin inhibitor. The  $\alpha$ -keto thiazole inhibitor **9** exhibited a 40-fold increase in potency to inhibit thrombin as compared to argatroban.




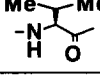
**Table 2. Effect of modifications in the P<sub>1</sub> position on thrombin inhibitors**

P<sub>3</sub>    P<sub>2</sub>    P<sub>1</sub>    P<sub>1'</sub>

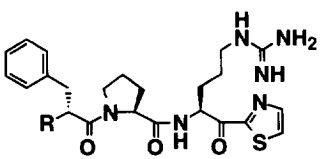
No	R	Inhibitory Activity IC <sub>50</sub> (μM)		Selectivity Trypsin/Thrombin
		Thrombin	Trypsin	
<b>9</b>	-(CH <sub>2</sub> ) <sub>3</sub> -N <sup>H</sup> H NH <sub>2</sub>	0.0015	0.0042	2.8
<b>20</b>	-(CH <sub>2</sub> ) <sub>4</sub> -NH <sub>2</sub>	0.26	1.2	4.6
<b>21</b>	-(CH <sub>2</sub> ) <sub>5</sub> -NH <sub>2</sub>	14	16	1.1
<b>Argatroban</b>		0.060	not determined	

The proline residue at the P<sub>2</sub> position in **9** was replaced by azetidine-2-carboxylic acid **10**, pipercolinic acid **11**, or valine **12** in order to investigate the influence of conformationally constrained amino acid (Table 3). While the azetidine-2-carboxylic acid derivative **10** maintained potency for the inhibition of thrombin, the pipercolinic acid derivative **11** was 480-times less active than **9**. Compound **10** exhibited the same selectivity for thrombin over trypsin as **9**. These results coincided with the results of Shuman et al.<sup>15</sup> Replacement of proline with valine did not significantly change the potency for the inhibition of thrombin, although the valine derivative **12** exhibited remarkable selectivity for thrombin over trypsin (trypsin/thrombin=21). This selectivity would be due to a conformational difference between the S<sub>2</sub> subsite of thrombin and trypsin.

**Table 3. Effect of modifications in the P<sub>2</sub> position on thrombin inhibitors**


No	A	Inhibitory Activity IC <sub>50</sub> (μM)		Selectivity Trypsin/Thrombin
		Thrombin	Trypsin	
10		0.003	0.009	3.0
9		0.0015	0.0042	2.8
11		0.73	41	56
12		0.0086	0.18	21

The substitution of the D-phenylalanine residue in **9** changed the inhibitory potency of thrombin (Table 4). Deletion of an amino group at the P<sub>3</sub> position showed the marked decrease in potency (**9** vs **13**). The importance of amino group was further investigated by acylation of compound **9**. A difference in inhibitory activity between the acetyl and benzyloxycarbonyl(Cbz) derivatives (**14** vs **15**) was observed. The acetyl compound **14** exhibited a 1000-fold loss in potency for the inhibition of thrombin as compared to compound **15**. The addition of the acetyl group in **9** led to a 20-fold decrease in selectivity. In this study, the modification at the P<sub>3</sub> position demonstrated no improvement in selectivity for thrombin over trypsin. On the other hand, the aldehyde derivative incorporating lactam sulfonamide moiety are reported to display potent and selective inhibitor.<sup>16</sup> The sulfonamide at the P<sub>3</sub> position might be a useful moiety for thrombin inhibitor.

**Table 4. Effect of modifications in the P<sub>3</sub> position on thrombin inhibitors**


No	R	Inhibitory Activity IC <sub>50</sub> (μM)		Selectivity Trypsin/Thrombin
		Thrombin	Trypsin	
13	-H	0.82	0.23	0.3
9	-NH <sub>2</sub>	0.0015	0.0042	2.8
14	-NHAc	2.3	0.23	0.1
15	-NHCbz	0.002	0.0016	0.8

Recently, Costanzo et al. reported that  $\alpha$ -keto benzothiazole inhibitor displays selectivity for thrombin compared with trypsin or plasmin.<sup>17</sup> The moiety of  $\alpha$ -keto benzothiazole in the P<sub>1'</sub> position would also provide a novel interaction with thrombin, but not trypsin.

The compound **9** with D-Phe-Pro-Arg motif showed the most potent inhibitory activity for thrombin (IC<sub>50</sub> value: 1.5 nM). IC<sub>50</sub> values ( $\mu$ M) for other serine proteases were as follows: trypsin, 0.0042; urokinase, 15.3; plasmin, >100; plasma kallikrein, >100.<sup>12</sup> These results suggested that the selectivity for thrombin over other proteases could be attributed to the different recognition of a D-Phe-Pro-Arg motif.

Finally, we evaluated the antithrombotic potency of compound **9** in comparison with argatroban for the ability to prolong prothrombin time (PT) and activated partial thromboplastin time (APTT) in rats (Table 5). Five minutes after intravenous administration of compound **9**, PT was prolonged 1.5 fold over the control value at 1 mg/kg and 12 fold at 3 mg/kg. A similar inhibitory potency was observed following intravenous administration of 1 mg/kg and 3 mg/kg with 2.0 fold and 8.3 fold APTT elevation, respectively. Although the  $\alpha$ -keto thiazole compound **9** showed a 40-fold increase in potency to inhibit the amidolytic activity of thrombin as compared to argatroban, the antithrombotic activity of compound **9** was about 2-fold more potent than that of argatroban. A possible explanation for the decreased activity of compound **9** in coagulation assays is that  $\alpha$ -keto thiazole compound might be a slow binding inhibitor and/or was less stable in vivo than argatroban.

**Table 5. Antithrombotic effect of compound **9** on intravenous administration**

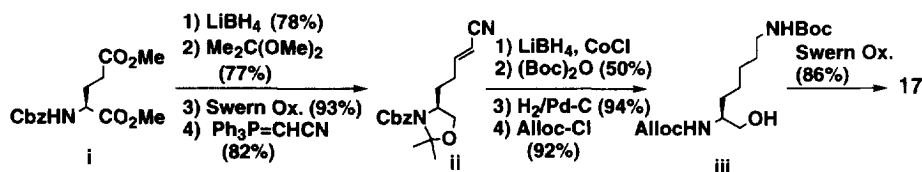
Compound	Dose (mg/kg)	PT inhibitor / PT control	APPT inhibitor / APPT control
<b>9</b>	1	1.5	2.0
<b>9</b>	3	12	8.3
Argatroban	3	6.2	4.8

**Conclusion:** We extended the utility of  $\alpha$ -keto heterocyclic compound to thrombin inhibitors. Tripeptidyl  $\alpha$ -keto thiazoles exhibited the inhibitory activity of thrombin at nanomolar levels. The modification at the P<sub>2</sub> position altered the selectivity for thrombin over trypsin. Compound **9** exhibited a more potent prolongation effect on the clotting time than argatroban at a dose of 3mg/kg intravenously. The tripeptide derivative with  $\alpha$ -keto heterocycle represented a promising new thrombin inhibitor.

## References and Notes

1. Stubbs, M. T.; Bode, W. *Trends. Biochem. Sci.* **1995**, 20, 23-28.
2. Tapparelli, C.; Metternich, R.; Ehrhardt, C.; Cook, N. S.; *Trends. Pharmacol. Sci.* **1993**, 14, 366-376. Stone, S. R. *Trends. Cardiovasc. Med.* **1996**, 5, 134-140. Edmunds, J. J.; Rapundalo, S. T. *Annu. Rep. Med. Chem.* **1996**, 31, 51-60.
3. Wiley, R. A.; Rich, D. H. *Med. Res. Rev.* **1993**, 13, 327-384.
4. Edwards, P. D.; Meyer, E. F., Jr.; Vijayalakshmi, J.; Tuthill, P. A.; Andisik, D. A.; Gomes, B.; Strimpler, A. *J. Am. Chem. Soc.* **1992**, 114, 1854-1863.

5. Tsutsumi, S.; Okonogi, T.; Shibahara, S.; Patchett, A. A.; Christensen, B. G.;  
*Bioorg. Med. Chem. Lett.* **1994**, 4, 831-834.  
Tsutsumi, S.; Okonogi, T.; Shibahara, S.; Ohuchi, S.; Hatsushiba, E.; Patchett, A. A.;  
Christensen, B. G. *J. Med. Chem.* **1994**, 37, 3492-3502.
6. Meiji Seika Kaisha, Ltd. JP08020597-A
7. We prepared **17** from the glutamic acid ester **i** by using the methods shown below.



8. Kawabata, S.; Miura, T.; Morita, T.; Kato, H.; Fujikawa, K.; Iwanaga, S.; Takada, K.; Kimura, T.; Sakakibara, S. *Eur. J. Biochem.* **1988**, 172, 17-25.
9. Walter, R.; Simmons, W. H.; Yoshimoto, T. *Molecular & Cellular Biochemistry* **1980**, 30, 111-127.
10. Bieth, J.; Spiess, B.; Wermuth, C. G. *Biochemical Medicine* **1974**, 11, 350-357.
11. Morita, T.; Kato, H.; Iwanaga, S.; Takada, K.; Kimura, T.; Sakakibara, S.  
*J. Biochem.* **1977**, 82, 1495-1498.
12. Kikumoto, R.; Tamao, Y.; Tezuka, T.; Tonomura, S.; Hara, H.; Ninomiya, K.; Hijikata, A.; Okamoto, S. *Biochemistry*. **1984**, 23, 85-90.
13. Takamiya, O. *Modern Medical Laboratory* **1991**, 19, 187-190.
14. Suzuki, S. *Modern Medical Laboratory* **1991**, 19, 191-195.
15. Shuman, R. T.; Rothenberger, R. B.; Campbell, C. S.; Smith, G. F.; Gifford-Moore, D. S.; Paschal, J. W.; Gesellchen, P. D. *J. Med. Chem.* **1995**, 38, 4446-4453.
16. Semple, J. E.; Rowley, D. C.; Brunck, T. K.; Ha-Uong, T.; Minami, N. K.; Owens, T. D.; Tamura, S. Y.; Goldman, E. A.; Siev, D. V.; Ardecky, R. J.; Carpenter, S. H.; e, Y.; Richard, B. M.; Nolan, T. G.; Hakanson, K.; Tulinsky, A.; Nutt, R. F.; Ripka, W. C. *J. Med. Chem.* **1996**, 39, 4531-4536.
17. Costanzo, M. J.; Maryanoff, B. E.; Hecker, L. R.; Schott, M. R.; Yabut, S. C.; Zhang, H.-C.; Andrade-Gordon, R.; Kauffman, J. A.; Lewis, J. M.; Krishnan, R.; Tulinsky, A.  
*J. Med. Chem.* **1996**, 39, 3039-3043.

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